

COLLINS Stephan-Christopher

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Email: stephanocollins@gmail.com

Nationality:

English / French

Date of birth:

10/09/74

Professional experience

Permanent academic position

GAD (C. Thauvin) – groupe NeuroGeMM (B. Yalcin)

Université de Bourgogne, 6, Bd Gabriel, 21000 Dijon, France

Maître de conférence (lecturer)

40% researcher at IGBMC, Strasbourg, since Sep 2015

September
2012-until
now

Project Manager (B. Thorens)

Center for Integrative Genomics Physiology Department,

UNIL, Genopode Building CH-1015 Lausanne

Researcher

September
2011-August
2012

Postdoctoral experience (P. Rorsman)

Oxford Centre for Diabetes, Endocrinology and Metabolism, Oxford University

Churchill hospital, Headington, OX3 7LJ, Oxford. UK

Research Scientist – Cell biology

September
2005-August
2011

Genetics Unit, Biochemistry Dpt, Oxford University (A. Russ)

South Parks Rd, OX1 3QU, Oxford. UK

Research Scientist – GPCR phylogenetic analysis

January 2005
– June 2005

The Wellcome Trust Sanger Institute (I. Barroso)

Hinxton, CB10 1SA Cambridge. UK

Research Scientist - Genetics

April 2003 –
December
2004

PhD (D. Gauguier)

The Wellcome Trust Centre for Human Genetics, Oxford University

Roosevelt Drive, Headington, OX3 7BN Oxford. UK

- D Phil

Subject: Rat chromosome 1 as a paradigm of human metabolic disorders

March 2000 –
March 2003 –

Temporary under- and post-graduate positions and national service

Research Assistant, The Wellcome Trust Centre for Human Genetics, Oxford University

Roosevelt Drive, Headington, OX3 7BN Oxford. UK

June 1998 –
March 2000

Laboratoire de Biochimie et Toxicologie Cliniques, HIA S^{te} Anne

BVD S^{te} Anne, 83 800 Toulon naval. France

Medical Laboratory Scientific Officer (*French National Service*)

October 1997
– May 1998

Laboratoire de physiopathologie de la nutrition, CNRS ESA 7059, Université Paris 7 (C. Magnan)

2 Place Jussieu, 75251 Paris cedex 05. France

Mars 1997 –
September
1997

Peer reviewed publications (total: 44, research gate score: 35.87, h-index 27)

Protocols

1. *Curr Protoc Mouse Biol.* 2018 Jun 26

A Method for Parasagittal Sectioning for Neuroanatomical Quantification of Brain Structures in the Adult Mouse

Collins SC, Kretz P, Wagner C, Kannan M, Fischer MC, Kessler P, Yalcin B

Genetics and bio-informatics

2. *Am J Hum Genet.* 2020 Nov 16;S0002-9297(20)30398-0. doi: 10.1016/j.ajhg.2020.11.001. Online ahead of print.

Heterozygous Variants in KDM4B Lead to Global Developmental Delay and Neuroanatomical Defects.

Duncan AR, Vitobello A, **Collins SC**, Vancollie VE, Lelliott CJ, Rodan L, Shi J, Seman AR, Agolini E, Novelli A, Prontera P, Guillen Sacoto MJ, Santiago-Sim T, Trimouille A, Goizet C, Nizon M, Bruel AL, Philippe C, Grant PE, Wojcik MH, Stoler J, Genetti CA, van Dooren MF, Maas SM, Alders M, Faivre L, Sorlin A, Yoon G, Yalcin B, Agrawal PB.

3. *Epilepsia.* 2020 May;61(5):868-878. doi: 10.1111/epi.16494. Epub 2020 Apr 2.

A knock-in mouse model for KCNQ2-related epileptic encephalopathy displays spontaneous generalized seizures and cognitive impairment.

Milh M, Roubertoux P, Biba N, Chavany J, Spiga Ghata A, Fulachier C, **Collins SC**, Wagner C, Roux JC, Yalcin B, Félix MS, Molinari F, Lenck-Santini PP, Villard L.

4. *J Anat.* 2019 Sep;235(3):637-650. doi: 10.1111/joa.13013. Epub 2019 Jun 7.

The neuroanatomy of Eml1 knockout mice, a model of subcortical heterotopia.

Collins SC, Uzquiano A, Selloum M, Wendling O, Gaborit M, Osipenko M, Birling MC, Yalcin B, Francis F.

5. *Nat Commun.* 2019 May 13;10(1):2129. doi: 10.1038/s41467-019-10081-8.

TUBG1 missense variants underlying cortical malformations disrupt neuronal locomotion and microtubule dynamics but not neurogenesis.

Ivanova EL, Gilet JG, Sulimenko V, Duchon A, Rudolf G, Runge K, **Collins SC**, Asselin L, Broix L, Drouot N, Tilly P, Nusbaum P, Vincent A, Magnant W, Skory V, Birling MC, Pavlovic G, Godin JD, Yalcin B, Héroult Y, Dráber P, Chelly J, Hinckelmann MV.

6. *Nat Commun.* 2019 Aug 1;10(1):3465

Large-scale neuroanatomical study uncovers 198 gene associations in mouse brain morphogenesis.

Collins SC, [...] , Moine H, Adams D, Reymond A, Lelliott CJ, Caleb Webber C, Yalcin B

7. *J Anat.* 2019 Sep;235(3):637-650

The neuroanatomy of Eml1 knockout mice, a model of subcortical heterotopia.

Collins SC, Uzquiano A, Selloum M, Wendling O, Gaborit M, Osipenko M, Birling MC, Yalcin B, Francis F.

8. *Nat Commun.* 2019 May 13;10(1):2129

TUBG1 missense variants underlying cortical malformations disrupt neuronal locomotion and microtubule dynamics but not neurogenesis.

Ivanova EL, Gilet JG, Sulimenko V, Duchon A, Rudolf G, Runge K, **Collins SC**, Asselin L, Broix L, Drouot N, Tilly P, Nusbaum P, Vincent A, Magnant W, Skory V, Birling MC, Pavlovic G, Godin JD, Yalcin B, Héroult Y, Dráber P, Chelly J, Hinckelmann MV.

9. *Genomics.* 2018 Mar;110(2):98-111

Molecular genetics of the transcription factor GLIS3 identifies its dual function in beta cells and neurons.

Calderari S, Ria M, Gérard C, Nogueira TC, Villate O, **Collins SC**, Neil H, Gervasi N, Hue C, Suarez-Zamorano N, Prado C, Cnop M, Bihoreau MT, Kaisaki PJ, Cazier JB, Julier C, Lathrop M, Werner M, Eizirik DL, Gauguier D

10. *Proc Natl Acad Sci U S A.* 2017 Oct 31;114(44):E9308-E9317

WD40-repeat 47, a microtubule-associated protein, is essential for brain development and autophagy.

Kannan M, Bayam E, Wagner C, Rinaldi B, Kretz PF, Tilly P, Roos M, McGillewie L, Bär S, Minocha S, Chevalier C, Po C; Sanger Mouse Genetics Project, Chelly J, Mandel JL, Borgatti R, Piton A, Kinnear C, Loos B, Adams DJ, Héroult Y, **Collins SC**, Friant S, Godin JD, Yalcin B.

11. *Am J Hum Genet.* 2017 Oct 5;101(4):564-577

The Immune Signaling Adaptor LAT Contributes to the Neuroanatomical Phenotype of 16p11.2 BP2-BP3 CNVs.
Loviglio MN, Arbogast T, Jøneh AE, **Collins SC**, Popadin K, Bonnet CS, Giannuzzi G, Maillard AM, Jacquemont S;
16p11.2 Consortium, Yalcin B, Katsanis N, Golzio C, Raymond A.

12. *Genome Med.* 2016 Sep 30;8(1):101.

Topological analysis of metabolic networks integrating co-segregating transcriptomes and metabolomes in type 2 diabetic rat congenic series.

Dumas ME, Domange C, Calderari S, Martínez AR, Ayala R, Wilder SP, Suárez-Zamorano N, **Collins SC**, Wallis RH, Gu Q, Wang Y, Hue C, Otto GW, Argoud K, Navratil V, Mitchell SC, Lindon JC, Holmes E, Cazier JB, Nicholson JK, Gauguier D.

13. *G3: Genes|Genomes|Genetics.* 2016 Sept.

Transcriptome profiling in rat inbred strains and experimental cross reveals discrepant genetic architecture of genome-wide gene expression.

Kaisaki PJ, Otto GW, Argoud K, **Collins SC**, Wallis RH, Wilder SP, Yau ACW, Hue C, Calderari S, Bihoreau MT, Cazier JB, Mott R, Gauguier D

14. *Diabetologia.* 2009 Dec;10(6):455-65.

Global microRNA expression profiles in insulin target tissues in a spontaneous rat model of Type 2 diabetes.

Herrera B, Lockstone HE, Taylor JM, Ria M, Barrett A, **Collins SC**, Kaisaki K, Argoud K, Fernandez C, Travers ME, Grew JP, Randall JC, Gloyn AL, Gauguier D, McCarthy MI, Lindgren CM.

15. *N Engl J Med.* 2007 Jan 18;356(3):237-47

Clinical and Molecular Genetic Spectrum of Congenital Leptin Receptor Deficiency

Farooqi SI, Wagenstein T, **Collins SC**, Kimber W, Matarese G, Keogh JM, Lank E, Bottomley B, Lopez-Fernandez J, Ferraz-Amaro I, Dattani MT, Ercan O, Grethe Myhre A, Retterstol L, Stanhope R, Edge J, MacKenzie S, Lessan N, Ghodsi M, De Rosa V, Perna F, Fontana S, Barroso I, Undlien DE, O'Rahilly S

16. *Diabetologia.* 2007 Mar;50(3):555-62

Adiponectin receptor genes: mutation screening in syndromes of insulin resistance and association studies for type 2 diabetes and metabolic traits in UK populations

Collins SC, Luan J, Thompson AJ, Daly A, Semple RK, O'Rahilly S, Wareham NJ, Barroso I

17. *Mamm Genome.* 2006 Jun;17(6):538-47 § shared authorship.

Mapping diabetes QTL in an intercross derived from a congenic strain of the Brown Norway and Goto-Kakizaki rats.

Collins SC, Wallis RH, Wilder SP, Wallace KJ, Argoud K, Kaisaki PJ, Bihoreau MT & Gauguier D.

18. *Physiol. Genomics.* 2004 Sep 16;19(1): 1-10

Quantitative trait locus dissection in congenic strains of the Goto-Kakizaki rat identifies a region conserved with diabetes loci in human chromosome 1q

Collins SC, Wallace KJ, Wallis RH, Argoud K, Kaisaki PJ, Ktorza A, Woon PY, Bihoreau MT & Gauguier D.

19. *Diabetologia.* 2004 Jun;47(6):1096-106

Enhanced insulin secretion and cholesterol metabolism in congenic strains of the spontaneously diabetic (Type 2) Goto Kakizaki rat are controlled by independent genetic loci in rat chromosome 8

Wallis RH, Wallace KJ, **Collins SC**, McAteer M, Argoud K, Bihoreau MT, Kaisaki PJ, Gauguier D.

20. *Mamm Genome.* 2003 May;14(5):350-6

Marker Assisted Congenic Screening (MACS): A Database Tool for the Efficient Production and Characterisation of Congenic Lines

Collins SC, Wallis RH, Wallace K, McAteer M, Bihoreau MT, Gauguier D

21. *Genomics.* 2001; 75(1-3): 57-69

A high-resolution consensus linkage map of the rat, integrating radiation hybrid and genetic maps

Bihoreau MT, Sebag Montefiore L, Godfrey RF, Wallis R H, Brown J H, Danoy PA, **Collins SC**, Rouard M, Kaisaki PJ, Lathrop M, Gauguier D

22. *Genomics.* 2000; 64(1): 32-43

Detailed comparative gene map of rat chromosome 1 with mouse and human genomes and physical mapping of an evolutionary chromosomal breakpoint

Kaisaki PJ, Rouard M, Danoy P A, Wallis R H, **Collins SC**, Rice M, Levy ER, Lathrop M, Bihoreau MT, Gauguier D

Cellular/whole body functional studies

23. *Mol Metab.* 2019 Feb;20:166-177
Mitochondrial Dynamin-Related Protein 1 (DRP1) translocation in response to cerebral glucose is impaired in a rat model of early alteration in hypothalamic glucose sensing.
Desmoulins, [...] , **Collins SC**, [...], et al.
24. *Plos One* ; 2018 Mar 15 ; 13(3)
Modulation of large dense core vesicle insulin content mediates rhythmic hormone release from pancreatic beta cells
Quinault A, Leloup C, Denwood G, Spiegelhalter C, Rodriguez M, Lefebvre P, Messaddeq N, Zhang Q, Dacquet C, Pénicaud L, **Collins SC**
25. *Proc Natl Acad Sci U S A.* 2017 Oct 31;114(44):E9308-E9317
WD40-repeat 47, a microtubule-associated protein, is essential for brain development and autophagy.
Kannan M, Bayam E, Wagner C, Rinaldi B, Kretz PF, Tilly P, Roos M, McGillewie L, Bär S, Minocha S, Chevalier C, Po C; Sanger Mouse Genetics Project, Chelly J, Mandel JL, Borgatti R, Piton A, Kinnear C, Loos B, Adams DJ, Hérault Y, **Collins SC**, Friant S, Godin JD, Yalcin B.
26. *Am J Hum Genet.* 2017 Oct 5;101(4):564-577
The Immune Signaling Adaptor LAT Contributes to the Neuroanatomical Phenotype of 16p11.2 BP2-BP3 CNVs.
Loviglio MN, Arbogast T, Jøneh AE, **Collins SC**, Popadin K, Bonnet CS, Giannuzzi G, Maillard AM, Jacquemont S; 16p11.2 Consortium, Yalcin B, Katsanis N, Golzio C, Reymond A.
27. *Diabetes.* 2016 Jul ;65(7) :1952-61. *Auteur correspondant*
Increased expression of the diabetes gene SOX4 reduces insulin secretion by impaired fusion pore expansion.
Collins SC, Do H, Hastoy B, Hugill A, Adam J, Chibalina M, Galvanovskis J, Lee S, Goldsworthy M, Salehi A, Tarasov A, Rosengren A, Cox R, Rorsman P
28. *Neuroscience.* 2015 Jun 10. pii: S0306-4522(15)00538-2
Glucose and hypothalamic astrocytes: More than a fueling role?
Leloup C, Allard C, Carneiro L, Fioramonti X, **Collins SC**, Pénicaud L
29. *J Biol Chem.* 2015 Jul 7. pii: jbc.M115.671248
NAADP and endolysosomal two-pore channels modulate membrane excitability and stimulus-secretion coupling in mouse pancreatic β cells.
Arredouani A, Ruas M, **Collins SC**, Parkesh R, Clough F, Pillinger T, Coltart G, Rietdorf K, Royle A, Johnson P, Braun M, Zhang Q, Sones W, Shimomura K, Morgan AJ, Lewis AM, Chuang KT, Tunn R, Gadea J, Teboul L, Heister PM, Tynan PW, Bellomo EA, Rutter GA, Rorsman P, Churchill GC, Parrington J, Galione A
30. *Cell Metab.* 2013 Dec 3;18(6):871-82
Role of KATP channels in glucose-regulated glucagon secretion and impaired counterregulation in type 2 diabetes.
Zhang Q, Ramracheya R, Lahmann C, Tarasov A, Bengtsson M, Braha O, Braun M, Brereton M, **Collins SC**, Galvanovskis J, Gonzalez A, Groschner LN, Rorsman NJ, Salehi A, Travers ME, Walker JN, Gloyn AL, Gribble F, Johnson PR, Reimann F, Ashcroft FM, Rorsman P
31. *Neurosci Lett.* 2013 Feb 8;534:75-9
Alteration of hypothalamic glucose and lactate sensing in 48h hyperglycemic rats.
Allard C, Carneiro L, **Collins SC**, Chrétien C, Grall S, Pénicaud L, Leloup C.
32. *Diabetologia*, 2011 Dec; 22
Multivesicular exocytosis in rat pancreatic beta cells.
Hoppa M, Jones E, Karanauskite J, Ramracheya R, Braun M, **Collins SC**, Zhang Q, Clark A, Eliasson L, Genoud C, Macdonald PE, Montheith AG, Barg S, Galvanovskis J, Rorsman P
33. *Diabetes*, 2010 May;59(5):1192-201. *Auteur correspondant*
Progression of diet-induced diabetes in C57Bl6J mice involves functional dissociation of Ca^{2+} channels from secretory vesicles.
Collins SC, Hoppa MB, Walker JN, Amisten S, Abdulkhader F, Bengtsson M, Fearnside J, Ramracheya R, Toye A, Zhang Q, Clark A, Gauguier D, Rorsman P.
34. *Cell Metab.* 2009 Dec;10(6):455-65.
Chronic palmitate exposure inhibits insulin secretion by dissociation of Ca^{2+} channels from secretory granules.
Hoppa MB, **Collins SC**, Ramracheya R, Hodson L, Amisten S, Zhang Q, Johnson P, Ashcroft FM, Rorsman P.
35. *Cell Metab.* 2009 Oct;10(4):309-15.

Suppression of sulfonylurea- and glucose-induced insulin secretion in vitro and in vivo in mice lacking the chloride transport protein CIC-3.

Li DQ, Jing X, Salehi A, **Collins SC**, Hoppa MB, Rosengren AH, Zhang E, Lundquist I, Olofsson CS, Mörgelin M, Eliasson L, Rorsman P, Renström E.

36. *Proc Natl Acad Sci U S A*. 2009 Apr 7;106(14):5813-8.

miR-375 maintains normal pancreatic α - and β -cell mass.

Poy MN, Hausser J, Trajkovski M, Braun M, **Collins SC**, Rorsman P, Zavolan M, Stoffel M.

37. *J Clin Invest*. 2009 Jan;119(1):80-90.

Expression of an activating mutation in the gene encoding the KATP channel subunit Kir6.2 in mouse pancreatic beta cells recapitulates neonatal diabetes.

Girard CA, Wunderlich FT, Shimomura K, **Collins SC**, Kaizik S, Proks P, Abdulkader F, Clark A, Ball V, Zubcevic L, Bentley L, Clark R, Church C, Hugill A, Galvanovskis J, Cox R, Rorsman P, Brüning JC, Ashcroft FM

38. *Cell*. 2009 Jan 23;136(2):235-48.

Regulation of PKD by the MAPK p38 δ in insulin secretion and glucose homeostasis

Sumara G, Formentini I, **Collins SC**, Sumara I, Musialek R, Bodenmiller B, Ramracheya R, Caille D, Jiang H, Platt KA, Meda P, Aebersold R, Rorsman P, Ricci R

39. *Genes Dev*. 2008 Nov 15;22(22):3135-46. *§ equal contribution*

pVHL is a regulator of glucose metabolism and insulin secretion in pancreatic β -cells

Zehetner J, **Collins SC** §, Danzer C §, Eckhardt K, Gerber PA, Ballschmieter P, Galvanovskis J, Shimomura K, Ashcroft FM, Thorens B, Rorsman P, Krek W

40. *Diabetologia*. 2008 Sep;51(9):1689-93

Long-term exposure of mouse pancreatic islets to oleate or palmitate results in reduced glucose-induced somatostatin and oversecretion of glucagon.

Collins SC, Salehi A, Eliasson L, Olofsson CS, Rorsman P

41. *Plos one*. 2008 Aug 13;3(8): *§ shared authorship*

Pathophysiological, Genetic and Gene Expression Features of a Novel Rodent Model of the Cardio-Metabolic Syndrome

Collins SC §, Wallis RH §, Kaisaki PJ, Wallace KJ, Argoud K, Ria M, Ktorza A, Rorsman P, Bihoreau MT, Gauguier D

42. *Obesity* 2008 Mar;16(3):522-30.

Pancreatic ectopic fat is characterized by adipocyte infiltration and altered lipid composition.

Pinnick KE, **Collins SC**, Londos C, Gauguier D, Clark A, Fielding BA

43. *Diabetes*. 2007 Jul;56(7):1888-97

Long-term exposure to glucose and lipids inhibits glucose-induced insulin secretion downstream of granule fusion with plasma membrane.

Olofsson CS, **Collins SC**, Bengtsson M, Eliasson L, Salehi A, Shimomura K, Tarasov A, Holm C, Ashcroft F, Rorsman P

44. *J Clin Invest*, 1999 Feb;103(3): 413-419

Lipid infusion lowers sympathetic nervous activity and leads to increased β -cell responsiveness to glucose

Magnan C, **Collins SC**, Berthault M F, Kassis N, Vincent M, Gilbert M, Penicaud L, Ktorza A, Assimakopoulos – Jeannot F

Supervision

Graduated PhD students

2009-2011 D. Do, University of Oxford (obtained in 2012)

Graduated MSc students (since 2016)

2021 A. Brocard, University of Bourgogne Franche-Comté, France
2021 A. Lahmar, University of Bourgogne Franche-Comté, France
2021 Z. Allaoua, University of Bourgogne Franche-Comté, France
2020 M. Milhau, University of Bourgogne Franche-Comté, France
2020 B. Capi, University of Bourgogne Franche-Comté, France
2020 J. Roussey, University of Bourgogne Franche-Comté, France
2019 C. Montillot, University of Nice Sophia Antipolis, France (currently PhD)
2019 A. Da Costa, University of Bourgogne Franche-Comté, France
2018 M. Gaborit, University of Strasbourg, France (currently PhD)
2017 M. Kobler (Medical student), University of Strasbourg, France
2017 M. Meylan, University of Strasbourg, France (currently PhD)
2017 S. Ciscars-Velazquez, University of Strasbourg, France
2017 D. Gualberto, University of Strasbourg, France
2017 G. Leiningner, University of Strasbourg, France
2016 N. Demeure (Engineer), University of Strasbourg, France (currently PhD)
2016 J. Delevoye, University of Strasbourg, France
2016 L. Durieux, University of Strasbourg, France (currently PhD)

Graduated BSc students (since 2016)

2021 M. Dessolain, University of Bourgogne Franche-Comté, France
2020 G. Fleury, University of Bourgogne Franche-Comté, France
2020 G. Boursier, University of Bourgogne Franche-Comté, France
2019 N. Yahiaoui, University of Strasbourg, France
2018 C. Bonnet, University of Poitiers, France (currently PhD)
2018 P. Hahn, University of Strasbourg, France
2018 M. Ossipenko, University of Strasbourg, France
2017 S. Lottiaux, University of Strasbourg, France
2016 D. Huynh, University of Strasbourg, France
2016 C. Casana, University of Strasbourg, France
2016 E. Ramos-Morales, University of Strasbourg, France (currently PhD)

Graduated Intern students (since 2016)

2020 N. Kenani, University of Paris 18th, France
2019 E. Isturiz, Supbiotech Paris, France
2019 N. Pigeonneau, University of Strasbourg, France
2017 E. Aguilar, University of Strasbourg, France
2016 M. Litt, University of Strasbourg, France
2016 S. Ott, University of Strasbourg, France
2016 L. Gagliardi, University of Strasbourg, France

Visiting International ERASMUS students (since 2016)

2018 G. Evyapan, PhD student, Turkey
2018 E. Makedona, BSc student, Greece
2017 A. Pathak, BSc student, India
2017 E. Mehmetoglu, BSc student, Turkey
2016 D. Kumruoglu, BSc student, Turkey

Teaching (past and present)

192 Hours / year since 2012
Main topic: Animal biology and cellular biology
Main class year 1 and 2
90% of teachings in practical and “TD”

Lectures in Zoology

The tegument (year 1)
The digestive tract (year 1)
The central nervous system (year 1)
The peripheral nervous system (Year 1)

Lectures in Neurosciences

Communication nerveuse Détection cérébrale des nutriments et équilibre énergétique : cas du glucose (Masters)
Enjeux des Sciences du vivant (biologie de la cellule bêta pancréatique) (Masters)
Neurogénétique et développement du CNS (Masters)
Espèces actives de l’oxygène (EAOs) : toxicité cellulaire ou signalisation (Masters)

Lectures in Scientific English

Techniques in electrophysiology (License Pro and Masters)

Practical: year 1 and 2 in animal biology and cell biology. Responsible for year 1 zoology practical from 2016 to 2019

Invited conferences and lectures

2020 3rd The Allied Genetics Conference, Washington D.C., USA, Poster (online) –**co author**
2020 10th Assises de Génétique Humaine et Médicale, Tours, France, **Poster –co author**
2019 Club Neuro-génétique Nancy, **Talk –co author**
2019 3rd Meeting Gene Expression and Epigenome, Strasbourg, France, **Talk –co author**
2019 69th American Society of Human Genetics, Houston, USA, **Poster –co author**
2018 International Research Consortium on Corpus Callosum, California, USA, **Talk –co author**
2018 1st Advances in Neurodevelopmental Disorders, Strasbourg, France, **Poster –co author**
2018 Exocytosis & Endocytosis Subgroup Symposium **Talk –co author**
2017 31st International Mouse Genome Conference, Heidelberg, Germany, **Poster –co author**
2017 Advances in electrophysiology IGBMC, Illkirch **Talk**
2017 Advancing Personalised Medicine with Animal Models, Athens, Greece, **Talk –co author**
2016 Symposium : Circadian clocks and metabolic health (Strasbourg). Invitation Etienne Challet **Talk**
2014 Lecture IGBMC (Strasbourg). Invitation Gérard Gradwohl / Roméo Ricci. **Talk**
2010 Lecture INCI. Invitation Marie France Bader. **Talk**
2005 et 2009 : MRC (Medical Research Council), Harwell. lectures. **Talks**
2006 to 2010 : OCDEM, Oxford. **Talks**
2003 Sanger Institute **Talk**

Societies

2015-2016 : « Société Francophone de Chronobiologie (SFC)»
2014-2015 : « Société Francophone du Diabetes (SFD)»
2009 : « Biochemical Society »
2008-10 : Membre de « Diabetes UK »

Collaborators

2019-present Prof A. Crosby, University of Exeter Medical School, United Kingdom
2019-present Prof P. Agrawal, Boston Children's Hospital, United States
2018-present Dr F. Francis, University of Sorbonne, Paris, France
2018-present Prof E. Sherr, University of California San Francisco, United States
2017-present Prof C. Tyler-Smith, Wellcome Sanger Institute, Hinxton, United Kingdom
2017-present Prof M. Hurles, Wellcome Sanger Institute, Hinxton, United Kingdom
2016-present Prof C. Webber, Dementia Research Institute, Cardiff, United Kingdom
2016-present Prof J. Chelly, IGBMC, Strasbourg, France
2016-present Dr J. Godin, IGBMC, Strasbourg, France
2016-present Dr Y. Hérault, Mouse Clinical Institute, Strasbourg, France
2016-present Prof D. Adams, Wellcome Sanger Institute, Hinxton, United Kingdom

Education

Oxford University, UK

D. Phil.

Grant: Wellcome Prize Studentship

Nottingham Trent University, UK

BSc Honours 1997

Second Class, Division one

Institutional responsibilities

2020 Organizer of a Symposium on Brain Imaging, INSERM Dijon (50 participants)

2018 Organizer of a Symposium on Statistics, Strasbourg, France (50 participants)

2017 Organizer of a Symposium on Electrophysiology, IGBMC Strasbourg, France (50 participants)

Award

Oxford University Merit Award (2009)

Computing

Data acquisition and mining:

Pulse+pulsefit, Felix32, FileMaker Pro, Access, Excel, R

Data analysis:

SPSS, Origin, R

Manuscript preparation and Data presentation:

Word, Power Point, Corel draw, Adobe Illustrator, Affinity designer

Database management:

Advanced user of FileMaker pro

1. Marker Assisted Congenic Screening (*MACS*): A Database Tool For The Efficient Production And Characterisation Of Congenic Lines (see article)
2. Fully incorporated laboratory database used for administration of orders, grants, inventory, protocols, experiments, results and analysis.
3. A database designed for the archive of cell lines (requested by Prof Chris Pugh – Wellcome Trust)
4. Current database developer at GAD / NGMM

Other

Bilingual English / French

Full driving license